Effects of *Pimenta pseudocaryophyllus* extracts on gout: Anti-inflammatory activity and anti-hyperuricemic effect through xantine oxidase and uricosuric action

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**Abstract**

**Ethnopharmacological relevance:** Leaves infusion of *Pimenta pseudocaryophyllus* (Gomes) Landrum is used in Brazilian folk medicine to treat the predisposition to arthritical and gouty affections of the joints, fever and other diseases. A refreshing drink prepared with the specie is also used due to its diuretic, sedative and aphrodisiac actions.

**Aim of the study:** The study was undertaken to investigate the mechanisms of anti-hyperuricemic effect and anti-inflammatory activity of *P. pseudocaryophyllus* extracts.

**Materials and methods:** Anti-hyperuricemic effect was investigated using xanthine oxidase assay and uricosuric studies with rats in which hyperuricemia was induced by potassium oxonate and uric acid. Anti-inflammatory activity was investigated on MSU crystal-induced paw edema model. Ethyl acetate extracts of the leaves (EAL) and branches (EAB), ethanolic extracts of leaves (EEL) and branches (EEB) and aqueous extracts of leaves (AL) and branches (AB) were evaluated.

**Results:** The extracts of *P. pseudocaryophyllus* evaluated showed expressive results regarding the inhibition of xanthine oxidase enzyme *in vitro* and they were also able to reduce serum uric acid levels in hyperuricemic rats. The investigation of the mechanism of action, it was found that EAL, EAB, EEB, AB (125 and 250 mg/kg) and AL (250 mg/kg) promoted an increase on the urinary excretion of uric acid and EEL, EEB, AB (125 and 250 mg/kg) and EAB (250 mg/kg) were capable to inhibit liver xanthine oxidase. Treatments with EEL (125 and 250 mg/kg) and EEB (250 mg/kg) were able to reduce edema at 48th h. EAL and EAB (125 and 250 mg/kg) showed significant anti-inflammatory activity on monosodium urate crystal-induced paw edema model at all evaluated times.

**Conclusions:** The specie *P. pseudocaryophyllus* showed remarkable anti-hyperuricemic effects through uricosuric effects and inhibition of xantine oxidase and therefore can be considered as a promise in the treatment of diseases related to hyperuricemia. Moreover, ethyl acetate extracts had significant anti-inflammatory activity.

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Pimenta pseudocaryophyllus (Gomes) Landrum, popularly known as "catáia", "cravo-preto" or "louro-cravo" is a species from Myrtaceae family largely distributed in pantropical and subtropical regions (Fajemiroye et al., 2012; Paula et al., 2012). In folk medicine, the leaves of *P. pseudocaryophyllus* are used to prepare a refreshing drink known for its diuretic, sedative and aphrodisiac actions (Paula et al., 2012). The population of Guaraqueçaba, state of Paraná, Brazil, uses *P. pseudocaryophyllus* leaves infusion in the form of tea to treat the predisposition to arthritic and gouty affections of the joints, fever and other diseases (D’Angelis and Negrelle, 2014).

Previous studies about *P. pseudocaryophyllus* described its anti-inflammatory and sedative action (Fajemiroye et al., 2012) and anti-oxidative activity (Campanini et al., 2014).

In the present study, it was reported that *P. pseudocaryophyllus* extracts are capable to reduce serum uric acid levels in hyperuricemic rats. Thus, the mechanisms on which the substances contained in the specie extracts exert the anti-hyperuricemic effect were investigated. In a complementary way, the anti-inflammatory activity was also evaluated through an experimental model of gouty arthritis induced by monosodium urate (MSU) crystals.

2. Materials and methods

2.1. Chemicals and reagents

Xanthine, potassium oxonate, uric acid, probenecid, benz-bromarone, allopurinol and indomethacin were purchased from Sigma-Aldrich (USA). Ketamine and xylazine was obtained from Sespro Indústria e Comercio Ltda (Brazil). Uric acid assay kit was purchased from Bioclin (Brazil). Monosodium urate (MSU) crystals were prepared according to previously described method (Rasool and Varalakshmi, 2006). All other chemicals were the highest analytic grade available.

2.2. Plant material

Leaves and branches from *P. pseudocaryophyllus* (Gomes) Landrum were collected in Lagoa Santa, Minas Gerais, Brazil, in October 2012. Coordinates of the location of the plant: 19°32’ 49.16” S/43°54’ 30.39” O. Voucher specimens were deposited in the Herbarium of Instituto de Ciências Exatas e Biológicas – UFOP, Ouro Preto, Brazil, reference number (OUPR 25900). Plant specie was collected with permission of Instituto Chico Mendes de Conservação da Biodiversidade – ICMBio/Sistema de Autorização e Informação em Biodiversidade-SISBIO (license No. 17021-8). The plant botanical identification was realized by Dr. Marcos Eduardo Guerra Sobral, Departamento de Ciências Naturais, Universidade Federal de São João Del-Rei (UFSJ).

2.3. Preparation of plant extracts

Leaves and branches were air-dried and pulverized separately. Extraction of leaves (1742.0 g) and branches (2896.0 g) was carried out by percolation method using hexane, ethyl acetate and ethanol in order of increasing polarity until exhaustion. Solvents were removed under reduced pressure at 40 °C using a rotatory evaporator (Buchi), yielded the following dry extracts: hexanic leaves (HL, 30.0 g), hexanic branches (HB, 3.8 g), ethyl acetate leaves (EAL, 74.0 g), ethyl acetate branches (EAB, 50.0 g), ethanolic leaves (EEL, 236.0 g) and ethanolic branches (EEB, 182.0 g). The aqueous extracts were obtained by exhaustion percolation of 70.0 g of leaves and branches powder with distillated water. The water was removed by lyophilization, yielding 9.6 g of aqueous crude extract of leaves (AL) and 4.9 g of aqueous crude extract of branches (AB).

2.4. Preliminary phytochemical screening

The presence of tannins and polyphenols, flavonoids, glycosides and aglycones anthraquinones, triterpenes, steroids, coumarins, alkaloids, saponins, glycosides cardiotonic, and proanthocyanidins were evaluated in the extracts of *P. pseudocaryophyllus*. Analyses were performed using the standard procedure previously described (Farnsworth, 1966; Matos, 1997). Triterpenes and steroids were analyzed using Lieberman–Burchard reaction. The presence of saponins was evaluated using froth test. Coumarins, quinones and anthraquinones were researched in the extracts using alkali reaction. Presence of alkaloids was evaluated by the Dragendorff reaction. Tannins were analyzed using ferric trichloride and potassium ferrocyanide. Flavonoids were detected by hydrochloric acid–magnesium reaction and TLC sprayed with NP/PEG reagent. Anthocyanins and catechins were assayed using hydrochloric acid and sodium hydroxide.

2.5. In vitro xanthine oxidase assay

The assay with *P. pseudocaryophyllus* extracts was carried out using a previous methodology described by Ferraz Filha et al. (2006), with modifications. EAL, EAB, EEL and EEB were solubilized in distilled water and DMSO (1%) to give final concentrations of 10.0; 20.0; 40.0; 50.0 and 100.0 μg/mL, in order to obtain the IC50 values. In a bucket containing a volume of 500 μL of each sample was added 1.125 mL of phosphate buffer (pH 7.4) and 187.5 μL X.O enzyme (0.28 U/mL). This system was incubated at 30 °C for 10 min. After that, 1.375 mL of xanthine substrate was added and immediately absorbances were obtained every minute for 10 min at 295 nm (Varian BIO-50). Allopurinol, a xanthine oxidase inhibitor, was used as positive control (10 μg/mL). The results were expressed as percentage of XO inhibition and calculated as: % inhibition = (1 – test inclination/blank inclination) × 100.

2.6. Animals

Male albino Swiss mice (25–30 g) and male Wistar rats (180–280 g) were supplied by Universidade Federal de Ouro Preto and all the experimental procedures were approved by its Ethical Committee under registration numbers: 2012/66 and 2014/31. Animals were divided into experimental groups (n = 6), housed in plastic boxes and maintained on a 12-h light/12-h dark cycle, at room temperature of 25 °C. They were given standard chow and water *ad libitum*. Experimental procedures were performed in accordance to the Guide for the Care and Use of Laboratory Animals, published by the US National Institute of Health (NIH Publication, revised in 1985).

2.7. Uricosuric activity in rats

Murugaiyah and Chan (2009) previously described the animal model used in the anti-hyperuricemic study. Rats received potassium oxonate (200 mg/kg, intraperitoneal) and uric acid (1 g/kg, by gavage) to become hyperuricemic. Food and water were withdrawn overnight prior to the study. EAL, EAB, EEL and EEB (125 and 250 mg/kg) were prepared in 5% DMSO oil solution and the clinically used drugs, benz bromarone (10 mg/kg) and probenecid (50 mg/kg) were prepared in a mixture of 10% ethanol in 20% Tween 20 aqueous solution. The formulations were administered intraperitoneally to the rats 30 min after hyperuricemia induction. For normal and hyperuricemic controls, the animals were treated only with the vehicle. Then, the animals were placed in metabolic
cages with 100 mL of tap water. The urine collected in graduated tubes and the water intake were measured for 5 h after the treatments. Finally, the animals were anesthetized with an association of ketamine and xylasine (40 and 87 mg/kg, respectively), administered intraperitoneally, in order to collect the blood from abdominal aorta. Blood samples were maintained at room temperature until blood coagulation and, afterwards, the serum was obtained after centrifugation at 3000 g for 10 min. The serum and urine samples were stored at −20 °C until uric acid quantification, which was performed by colorimetric method, using a standard diagnostic kit (Bioclin, Brazil), according to manufacturer’s instructions.

2.8. In vivo xanthine oxidase assay

Rat’s livers were excised immediately after blood collection, washed in 0.9% saline and rapidly stored at −80 °C until processing. One group of animals was treated with allopurinol (10 mg/kg), known xanthine oxidase inhibitor, for positive control of this assay. The separation of cytosolic fraction containing the enzyme was performed as described elsewhere (Haidari et al., 2009; Zhu et al., 2004). Briefly, livers were homogenized in 5 mL of 80 mM sodium phosphate buffer (pH 7.4). The homogenate was centrifuged at 3000 g for 10 min at 4 °C. The lipid layer was removed and the supernatant was centrifuged again at 10,000 g for 60 min at 4 °C, resulting in the cytosolic fraction. This fraction was used to evaluate liver xanthine oxidase residual activity according to a method previously described by Hall et al. (1990) with modifications. The formation of uric acid was monitored spectrophotometrically. Briefly, 100 μL of the liver cytosolic fraction was pre-incubated in 5.4 mL of potassium oxonate solution (1 mM) in 50 mM sodium phosphate buffer (pH 7.4) at 35 °C for 15 min. After the incubation period, 1.2 mL of xanthine solution (250 mM) was added to start the reaction, which, in turn, was stopped after 0 and 30 min by the addition of 500 μL of 0.6 M HCl. The samples were centrifuged at 3000 g for 5 min and the uric acid in the supernatant was spectrophotometrically quantified at 295 nm (Varian BIO-50). Protein concentration was determined according Bradford (1976) method using bovine serum albumin as standard. Enzyme activity was expressed as nmoles of uric acid produced per min by 1 mg of protein [nmol/min mg protein].

2.9. Effects on monosodium urate crystal-induced inflammation in mice

Anti-inflammatory activity of *P. pseudocaryophyllus* extracts was evaluated using an animal model of paw edema adapted from the previously described method by Rasool and Varalakshmi (2006). On the first day of the experiment (time 0), the inflammation was induced by injection of 50 μL (80 mg/mL) of urate crystals suspension (MSU) into the subplantar region of the mouse right hind paw, while the normal control group received saline 0.9%. EAL, EAB, EEL and EEB (125 and 250 mg/kg) were prepared in 5% DMSO oily solution. Indomethacin (3 mg/kg) was prepared in a mixture of 10% Tween 20 aqueous solution. The samples were administered by gavage 1 h before MSU injection and repeated daily for 2 more days. For normal and inflammation controls, only vehicle was given to the animals. The paw thickness between the dorsal and ventral faces of the paws was obtained using a caliper rule at 0, 4, 24 and 48 h after the MSU injection, while inflammatory swelling was expressed as thickness variation (Δ) versus time 0.

2.10. Statistical analysis

The results were presented as mean ± standard error of the mean (S.E.M.) of six animals. The statistical significance of the difference was evaluated by analysis of variance (ANOVA) followed by Dunnett’s test using GraphPad Prism 5.0 Software (Inc., San Diego, CA, USA). The IC50 values were calculated by linear regression.

3. Results

Gout is an acute inflammatory arthritis caused by deposition of uric acid in the joints and tissues. About 10% of individuals with hyperuricemia develop gout and 90% of gout patients have hyperuricemia (Smith et al., 2010). The therapeutic control of gout consists in the treatment of the acute inflammatory process and reduction of blood uric acid levels. Gout management is based on the use of anti-inflammatory drugs, xanthine oxidase inhibitors and consequently inhibition of uric acid synthesis and drugs that increase the excretion of uric acid. In the present study, *in vitro* and *in vivo* xanthine oxidase inhibitory activity, anti-hyperuricemic, uricosuric and anti-inflammatory effects of extracts from *P. pseudocaryophyllus* were evaluated.

3.1. Screening of chemical compounds in the species

In pharmacognotic analyses were detected the presences of triterpenes, steroids, coumarins, saponins, tannins and flavonoids. The positive reaction of EEL, EEB, AL and AB with sodium nitrite, indicated the presence of hydrolysable tannins and the negative reaction with n-butanol, suggested the absence of condensed tannins in these extracts. Chromatographic plate containing samples of these extracts revealed with sulfuric acid confirmed the absence of proanthocyanidins. In EAB and EEL were identified the presence of coumarins. Flavonoids were detected in EAL, EAB, EEL, AL and AB. In aqueous extracts (AL and AB) were identified the presence of saponins. Triterpenes and steroids were detected in EAL.

3.2. In vitro xanthine oxidase inhibitory activity

The inhibitory activity of *P. pseudocaryophyllus* extracts against xanthine oxidase and their respective IC50 values are shown in Table 1. Allopurinol, positive control for the bioassay, showed an IC50 of 0.33 μg/mL. EAL, EAB and EEB showed IC50 values of 14.07; 15.57 and 13.73 μg/mL, respectively.

3.3. Anti-hyperuricemic effects of *Pimenta pseudocaryophyllus* extracts in hyperuricemic rats

Potassium oxonate solution significantly increased serum urate levels compared to normal control group (Table 2). Animals treated with extracts showed a reduction on serum uric acid levels, especially those animals treated with EAB (250 mg/kg). This

<table>
<thead>
<tr>
<th>Sample</th>
<th>Concentration (μg/mL)</th>
<th>Inhibition (%)</th>
<th>IC50 (μg/mL ± S.D.)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Allopurinol</td>
<td>10</td>
<td>95.63</td>
<td>0.33 ± 0.011</td>
</tr>
<tr>
<td>EAL</td>
<td>100</td>
<td>72.54</td>
<td>14.07 ± 1.689</td>
</tr>
<tr>
<td>EEL</td>
<td>100</td>
<td>76.47</td>
<td>19.91 ± 1.859</td>
</tr>
<tr>
<td>AL</td>
<td>100</td>
<td>45.94</td>
<td>109.7 ± 3.926</td>
</tr>
<tr>
<td>EAB</td>
<td>100</td>
<td>80.91</td>
<td>15.57 ± 1.561</td>
</tr>
<tr>
<td>EEB</td>
<td>100</td>
<td>81.63</td>
<td>13.72 ± 1.67</td>
</tr>
<tr>
<td>AB</td>
<td>100</td>
<td>57.05</td>
<td>54.65 ± 1.300</td>
</tr>
</tbody>
</table>

Ethyl acetate extract of leaves (EAL), ethanolic extract of leaves (EEL), aqueous extract of leaves (AL), ethyl acetate extract of branches (EAB), ethanolic extract of branches (EEB) and aqueous extract of branches (AB).
extract reduced uric acid to values that did not showed significant differences when compared to normal control group.

3.4. Uricosuric effects of *Pimenta pseudocaryophyllus* extracts in hyperuricemic rats

In general, treatment with *P. pseudocaryophyllus* extracts caused insignificant changes in water intake and urine output of hyperuricemic rats. These data, as well as the extracts effects on uric acid urinary excretion and serum uric acid are shown in Table 2. The rats became hyperuricemic by a single administration of potassium oxonate intraperitoneally (200 mg/kg) and uric acid orally (1 g/kg), showing a higher uric acid excretion when compared to normal control animals. Rats treated with clinically used drugs benz bromarone (10 mg/kg) and probenecid (50 mg/kg), as well as EAL, EAB, EEB, AB (125 and 250 mg/kg) and AL (250 mg/kg) promoted a significant increase in uric acid excretion when compared to hyperuricemic control animals.

3.5. In vivo xanthine oxidase inhibitory activity

Table 3 shows the effect of *P. pseudocaryophyllus* extracts on liver xanthine oxidase residual activity. Hyperuricemia induction did not cause any appreciable changes on liver xanthine oxidase residual activity, since this enzyme is not inhibited during the process of hyperuricaemia induction. A single dose of allopurinol (10 mg/kg) caused a significant reduction (76.14%) of liver xanthine oxidase residual activity. EEL, EEB, AB (125 and 250 mg/kg) and EAB (250 mg/kg) caused a significant inhibition of liver xanthine oxidase residual activity, especially EEL (125 and 250 mg/kg), that caused an enzyme inhibition of 45.55% and 45.16%, respectively.

3.6. Effects of *Pimenta pseudocaryophyllus* extracts on MSU-induced inflammation in mice

The edema observed on animals that received only vehicle after MSU crystals injection was considered as the maximum inflammation. It was used as reference and compared to other treatments in order to evaluate the anti-inflammatory activity (Table 4). Treatments with AL and AB were not able to reduce paw edema significantly at any tested dose. EEL (125 and 250 mg/kg) and EEB (250 mg/kg) significantly reduced paw edema at 48th h. Treatment with EAL and EAB were able to reduce edema at all evaluated times. Indomethacin (3 mg/kg) showed a significant anti-inflammatory activity similarly to that observed on groups treated with ethyl acetate and ethanol extracts from *P. pseudocaryophyllus*.
edema in mice.

Due to expressively inhibit xanthine oxidase 

The toxicity, whereas probenecid and sul

well as they should not be used in patients with history of ur-

Sparattosperma leucanthum

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A potential source of novel anti-hyperuricemic agents may be
derived from natural products. The methanol extract and lignans of Phyllanthus niruri showed anti-hyperuricemic effects, the meth-

administration of its uricosuric action and partly through xanthine oxidase inhibition, whereas the activity of the lignans was attributed to their uricosuric action (Murugaiyah and Chan, 2009). In previous studies, Lycnophora trichocarpha and Sparattosperma leucanthum proved to be a promise for the treat-

The levels of PGE2, α-amyrin, and β-amyrin were suppressed in these rats (Mor-

Further investigations (Schmeda-Hirschmann et al., 1996). As all extracts (except AL and AB) showed IC50 values under 50 μg/mL, they were in vivo evaluated.

Administration of P. pseudocaryophyllus extracts from leaves and branches caused a reduction in serum uric acid levels in hyperuricemic rats. This fact can be understood through the two pathways investigated. The extracts EAL, EAB, EEB and AB two evaluated doses and AL, (250 mg/kg) showed an increased urinary excretion of uric acid. It is estimated that 80–90% of patients with gout are under-

Inflammation is one of the major problems reported on gout patients and this relationship has been evaluated using in vitro studies, animal and human models (Krishnan, 2014). One of the main characteristics of acute gouty arthritis is the cell activation promoted by monosodium urate (MSU) microcrystals in joints, which can interact with cells such as neutrophils, monocytes, macrophages and synovial cells as fibroblasts (Jiang et al., 2012). For example, in monocytes, microcrystals stimulate the synthesis of a large number of pro-inflammatory cytokines such as tumor necrosis factor (TNF-α), IL-1β; IL-6 and IL-8 (Neogi, 2011). In addition, macrophages phagocyte urate crystals initiating the release of lysosomal enzymes involved in processes such as chemotaxis and cell permeability (Rasool and Varalakshmi, 2006). In the present study, it was observed that the ethyl acetate extracts (EAL and EAB) showed significant reduction in paw edema induced by 

This shows that these compounds can be partially attributed to

4. Discussion

Hyperuricemia is a metabolic disorder associated with the development of pathological conditions such as gout. The relationship between hyperuricemia and the pathogenesis of gout has been reported since the early nineteenth century (Neogi, 2011). Although the disease afflicts humans for years, there is a limited number of drugs currently used in clinical practice for treating hyperuricemia or gout, which are divided into two classes: xanthine oxidase inhibi-

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Previous phytochemical investigations of P. pseudocaryophyllus resulted in the isolation of the pentacyclic triterpenes lupeol, α-amyrin, and β-amyrin and the flavonoids quercetin, quercitrin, and azelein (Paula et al., 2012). Among these compounds, lupeol and quercetin have reports of reducing uric acid levels in hyperuricemic mice caused by oxonate. The hypouricemic effects of quercetin is partly due to inhibition of liver xanthine oxidase residual activity (Zhu et al., 2004). Lupeol does not have its hypouricemic mechanism described (De Souza et al., 2012).

These compounds are also related to anti-inflammatory activity. Quercetin also inhibited the inflammatory response induced by carrageenan in rats. The contents of PGE2, TNF-α, RANTES, MIP-2 and the mRNA for cyclooxygenase-2 were suppressed in these rats (Morikawa et al., 2003). Lupeol have inhibitory properties on the production of inflammatory cytokines as IL-1, TNF-α and PGE2 release by macrophages in vitro (Fernández et al., 2001). The pentacyclic tri-

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Table 4

<table>
<thead>
<tr>
<th>Group</th>
<th>Dose (mg/kg)</th>
<th>Thickness variation Δ (mm)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>4 h</td>
<td>24 h</td>
</tr>
<tr>
<td>Vehicle</td>
<td></td>
<td></td>
</tr>
<tr>
<td>(salina)</td>
<td>0.18 ± 0.038</td>
<td>0.16 ± 0.049</td>
</tr>
<tr>
<td>MU</td>
<td>1.72 ± 0.232</td>
<td>1.48 ± 0.200</td>
</tr>
<tr>
<td>Indomethacin</td>
<td>0.27 ± 0.046</td>
<td>0.43 ± 0.091</td>
</tr>
<tr>
<td>EAL</td>
<td>0.10 ± 0.041</td>
<td>0.11 ± 0.081</td>
</tr>
<tr>
<td>250</td>
<td>0.30 ± 0.095</td>
<td>0.48 ± 0.166</td>
</tr>
<tr>
<td>EEL</td>
<td>1.21 ± 0.209</td>
<td>0.98 ± 0.189</td>
</tr>
<tr>
<td>250</td>
<td>1.09 ± 0.277</td>
<td>1.04 ± 0.131</td>
</tr>
<tr>
<td>AL</td>
<td>1.17 ± 0.240</td>
<td>1.52 ± 0.154</td>
</tr>
<tr>
<td>250</td>
<td>1.33 ± 0.209</td>
<td>0.15 ± 0.185</td>
</tr>
<tr>
<td>EAB</td>
<td>0.20 ± 0.103</td>
<td>0.12 ± 0.030</td>
</tr>
<tr>
<td>250</td>
<td>0.27 ± 0.129</td>
<td>0.51 ± 0.131</td>
</tr>
<tr>
<td>EEB</td>
<td>1.63 ± 0.095</td>
<td>1.40 ± 0.265</td>
</tr>
<tr>
<td>250</td>
<td>1.12 ± 0.195</td>
<td>1.08 ± 0.222</td>
</tr>
<tr>
<td>AB</td>
<td>1.54 ± 0.078</td>
<td>1.32 ± 0.187</td>
</tr>
<tr>
<td>250</td>
<td>1.06 ± 0.079</td>
<td>1.15 ± 0.118</td>
</tr>
</tbody>
</table>
5. Conclusions

The specie P. pseudocaryophyllus showed remarkable anti-hyperuricemic activity. The tested extracts were able to reduce serum uric acid through the two main pathways of uric acid regulation in human body, by inhibiting its formation and increasing its excretion. Therefore, P. pseudocaryophyllus extracts can be considered promising in the treatment of diseases related to hyperuricemia. Moreover, ethyl acetate extracts also had significant anti-inflammatory activity.

Several flavonoids have shown the ability to inhibit xanthine oxidase, anti-hyperuricemic and anti-inflammatory activities. Flavonoids were detected in all evaluated extracts, being considered the predominant metabolites in P. pseudocaryophyllus. Therefore, this group of metabolites may be responsible for the anti-hyperuricemic and anti-inflammatory effects of P. pseudocaryophyllus extracts. However, only a full elucidation of the specie chemical profile will highlight the active substances.

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